

quenching ($k_q \sim 5 \times 10^8 \text{ s}^{-1} \text{ M}^{-1}$).^{15,16}

2. Neither a direct excitation of molecular oxygen ($^3\Sigma_g^- \text{O}_2$) nor a (O_2)₂ simultaneous transition is responsible for the $^1\Delta_g \text{O}_2$ signal under our conditions.^{10,17-20} Both cyclohexane and methanol can dissolve substantial amounts of oxygen²¹ and exhibit oxygen-dependent absorption bands to the blue (<260 nm) of our pump wavelengths, but neither yield a $^1\Delta_g \text{O}_2$ signal when irradiated at 355 or 266 nm. It is expected that discrete oxygen transitions should be solvent independent. Furthermore, a plot of $^1\Delta_g \text{O}_2$ signal intensity (I_Δ) versus oxygen ($^3\Sigma_g^- \text{O}_2$) concentration shows no positive curvature for any of the solvents mentioned above. This eliminates the possibility of a (O_2)₂ simultaneous absorption.

3. The I_Δ is linearly dependent on the excitation laser energy. This precludes a multiphoton absorption in the solvent and the subsequent formation of $^1\Delta_g \text{O}_2$ in a trivial sensitized process.

4. The excitation wavelengths used in our experiments are to the red of the solvent $S_1 \leftarrow S_0$ transition. Therefore, in a single photon absorption, the singlet excited electronic states of the solvent are energetically inaccessible.

5. The $^1\Delta_g \text{O}_2$ is not produced by an adventitious sensitizer either present as an impurity in the solvent or one which is photolytically created. Upon prolonged irradiation, I_Δ is constant, and the absorption spectra of the irradiated samples show no evidence of degradation. For each solvent, we are able to observe a small absorbance under nitrogen-purged conditions at the excitation wavelength using a 10 cm path length cell. In the case of mesitylene, *p*-xylene, *o*-xylene, and 1,4-dioxane, this absorbance can be reduced by successive distillation, indicating the presence of impurities in our samples. However, the reduction in the absorbance upon distillation is not accompanied by an equal reduction in I_Δ . To further show that these impurities are not responsible for $^1\Delta_g \text{O}_2$ formation, we prepared standard benzene solutions of a known $^1\Delta_g \text{O}_2$ sensitizer, acridine, such that the solution optical density (OD) at 355 nm was comparable to that of the impurity as measured in a nitrogen purged solvent. Acridine in benzene is an efficient $^1\Delta_g \text{O}_2$ sensitizer (quantum yield = 0.8).⁶ The I_Δ obtained by pumping the oxygen ($^3\Sigma_g^- \text{O}_2$) induced absorption bands in the aromatic solvents is at least three times greater than that observed for the standard acridine photosensitized $^1\Delta_g \text{O}_2$ signal. Although $^1\Delta_g \text{O}_2$ quantum yields of 2.0 are known, this phenomenon occurs at the limit of infinite oxygen concentration where all sensitizer singlet states are being quenched by $^3\Sigma_g^- \text{O}_2$.^{6,22} At the oxygen concentrations used in our experiments, the results presented above indicate that if the impurity is indeed responsible for $^1\Delta_g \text{O}_2$ formation, it must have a $^1\Delta_g \text{O}_2$ quantum yield which far exceeds that of a standard photosensitized process.⁶ Therefore, we conclude that $^1\Delta_g \text{O}_2$ is not produced by an adventitious sensitizer.

We suggest the observed $^1\Delta_g \text{O}_2$ will be formed primarily by a combination of two different channels: (1) a spin allowed, solvent-oxygen ($^3\Sigma_g^- \text{O}_2$) cooperative transition to a triplet charge transfer state¹ with subsequent intersystem crossing and dissociation to yield $^1\Delta_g \text{O}_2$ and the ground state solvent molecule and (2) an oxygen ($^3\Sigma_g^- \text{O}_2$) enhanced $T_1 \leftarrow S_0$ transition² followed by the sensitized formation of $^1\Delta_g \text{O}_2$ by the resulting triplet state. The wavelengths that correspond to the $T_1 \leftarrow S_0$ (0,0) absorption are as follows: benzene (340 nm),^{2,23} toluene (346 nm),²⁴ *o*-xylene

(348 nm),²⁴ *p*-xylene (355 nm),²⁴ and mesitylene (357 nm).²⁴ These distinct transitions are all found in the same spectral region as the broad, structureless Mulliken CT band. For benzene, toluene, and *o*-xylene, the excitation wavelength (355 nm)⁵ is too far to the red to excite the $T_1 \leftarrow S_0$ transition. If the enhanced $T_1 \leftarrow S_0$ transition were the only channel responsible for the formation of the observed $^1\Delta_g \text{O}_2$, we would expect to see the $^1\Delta_g \text{O}_2$ signal only when *p*-xylene and mesitylene are pumped, and that the signal from *p*-xylene might be more intense than that from mesitylene. This is not the case. Rather, the I_Δ is linearly dependent on the OD of the oxygen-induced absorption at 355 nm for oxygenated benzene, toluene, *o*-xylene, and mesitylene. The *p*-xylene point, for which the $T_1 \leftarrow S_0$ transition is coincident with the excitation wavelength (355 nm), falls above the line, but I_Δ is not as large as that for mesitylene. Therefore, although some of the $^1\Delta_g \text{O}_2$ produced may be attributed to an oxygen-induced $T_1 \leftarrow S_0$ transition, we suggest that $^1\Delta_g \text{O}_2$ is also produced upon dissociation of a photolytically generated CT state.

In conclusion, we have provided direct spectroscopic evidence for the formation of $^1\Delta_g \text{O}_2$ subsequent to UV irradiation of a solvent-oxygen cooperative absorption band.

Acknowledgment. We thank Maria P. Dillon and Kai-Kong Yu for their assistance at various stages of this project. This work was supported by the National Science Foundation under Grant Number CHE-8501542.

Use of $\text{Ir}_4(\text{CO})_{11}$ To Measure the Lengths of Organic Molecules with a Scanning Transmission Electron Microscope

Frederic R. Furuya, Larry L. Miller,* J. F. Hainfeld, William C. Christophel, and Peter W. Kenny

Department of Chemistry, University of Minnesota
Minneapolis, Minnesota 55455
Biology Department, Brookhaven National
Laboratory, Upton, New York 11973
Received September 21, 1987

We report an evaluation of the hypothesis that metal-cluster labels can be used to accurately measure molecular distances by transmission electron microscopy (TEM). Organic and most small biological molecules are essentially transparent in TEM, but distances between molecular sites can in principle be measured by attachment of electron-dense clusters to those sites. The present test utilized a sufficiently long and rigid organic spacer with specifically attached $\text{Ir}_4(\text{CO})_{11}$ labels and took advantage of the capabilities of scanning transmission electron microscopy (STEM) for high resolution measurements on molecular materials. $\text{Ir}_4(\text{CO})_{11}$ -labeled molecules, which have not previously been used for TEM, are shown to be synthetically accessible, sufficiently stable, and immobile in the electron beam so that they may have wider TEM applicability.

Electron microscopists have had a long standing interest in labeling molecular structures with metals. In order to take advantage of the high resolution of modern microscopes and achieve specificity in imaging, attention has been given to metal clusters. Two types of clusters have been studied: anionic heteropolytungstates,^{1,2} e.g., $(\text{RC}_5\text{H}_4)\text{TiP}_2\text{W}_{17}\text{O}_{61}^{7-}$, and cationic undecagold clusters,³ e.g., $\text{Au}_{11}(\text{CN})_3[\text{P}(\text{C}_6\text{H}_4\text{CH}_2\text{NH}_3^+)_3]_7$. Molecular

* Address correspondence to author at University of Minnesota.

(1) (a) Keana, J. F. W.; Ogan, M. D.; Lu, Y.; Beer, M.; Varkey, J. J. *Am. Chem. Soc.* **1985**, *107*, 6714. (b) Keana, J. F. W.; Ogan, M. D. *J. Am. Chem. Soc.* **1986**, *108*, 7951. (c) Keana, J. F. W.; Ogan, M. D.; Lu, Y.; Beer, M.; Varkey, J. J. *Am. Chem. Soc.* **1986**, *108*, 7956.

(2) Monson, K. L.; Wall, J. S.; Hainfeld, J. F. *Ultramicroscopy* **1987**, *21*, 147.

(3) (a) Safer, D.; Hainfeld, J. F.; Wall, J. S.; Riordan, J. *Science (Washington, D. C.)* **1982**, *218*, 290. (b) Frey, P. A.; et al. *Biochemistry* **1984**, *23*, 3849, 3857, 3863. (c) Safer, D.; Bolinger, L.; Leigh, J. S. *J. Inorg. Biochem.* **1986**, *26*, 77. (d) Lipka, J. J.; Hainfeld, J. F.; Wall, J. S. *J. Ultrastruct. Res.* **1983**, *84*, 120.

(15) Wilkinson, F.; Brummer, J. G. *J. Phys. Chem. Ref. Data* **1981**, *10*, 809-999.

(16) A small amount of water was added to the solution in order to dissolve NaN_3 .

(17) Rosenthal, I., in ref 10, pp 13-38 and references cited therein.

(18) (a) Eisenberg, W. C.; Taylor, K.; Veltman, J.; Murray, R. W. *J. Am. Chem. Soc.* **1982**, *104*, 1104-1105. (b) Eisenberg, W. C.; Snelson, A.; Butler, R.; Taylor, K.; Murray, R. W. *J. Photochem.* **1984**, *25*, 439-448.

(19) Koroll, G. W.; Singh, A. *Photochem. Photobiol.* **1978**, *28*, 611-613.

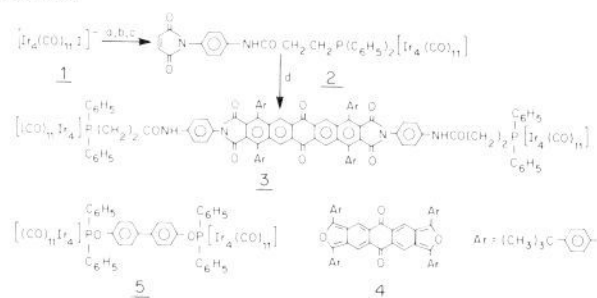
(20) Frei, H.; Pimentel, G. C. *J. Chem. Phys.* **1983**, *79*, 3307-3319.

(21) Battino, R., Ed. *IUPAC Solubility Data Series, Volume 7. Oxygen and Ozone*; Pergamon: Oxford, 1981; p 301.

(22) Dobrowolski, D. C.; Ogilby, P. R.; Foote, C. S. *J. Phys. Chem.* **1983**, *87*, 2261-2263.

(23) Sponer, H.; Kanda, Y.; Blackwell, L. A. *Spectrochim. Acta* **1960**, *16*, 1135-1147.

(24) Kanda, Y.; Shimada, R. *Spectrochim. Acta* **1961**, *17*, 279-285.

Scheme 1^a

^a (a) (C₆H₅)₂PCH₂CH₂CO₂H; THF, 24 h, room temperature (100%); (b) (COCl)₂, benzene/CHCl₃, 24 h, room temperature used without purification; (c) *N*-(*p*-aminophenyl)maleimide, CHCl₃, 2,6-di-*tert*-butylpyridine, 4 h, room temperature (79%); (d) **4**, CHCl₃, 18 h, room temperature, in the dark, followed by concentrated H₂SO₄ (28%).

separation distances have been measured between the four undecagold clusters which could be attached to avidin,^{3a} and appropriately paired spots were observed from an organic compound to which two heteropolytungstates were attached.^{1a} This observation was not analyzed in detail. Measurements of molecular distances are of interest to us for establishing the dimensions of molecular electronics structures, and it seemed important to more accurately test this TEM approach to distance measurement.

Et₄N⁺, [Ir₄(CO)₁₁I]⁻ (**1**), prepared⁴ from commercial Ir₄(CO)₁₂, was employed to demonstrate the TEM qualities of tetrairidium clusters. Initial studies employed a conventional transmission electron microscope (Hitachi H-600, 100 kV) in the bright field mode. These studies gave evidence for good stability of the cluster in the beam and were preparatory for more thorough high resolution studies. With use of the Brookhaven STEM⁵ which was operating at 40 kV, samples were subjected to a cumulative electron dose ranging from 5 × 10⁴ to 4 × 10⁵ e/nm². Specimen temperature was -135 °C during imaging. The sample was prepared by deposition from 10⁻⁶ M aqueous solution onto 25-Å carbon films. The dark field images showed bright spots ~6 Å in diameter. Computer analysis⁵ indicated a mass of 800 ± 200 (sd) as expected for a single cluster. The Kolmogorov-Smirnov test applied to histograms from nine micrographs indicated a *random distribution of nearest neighbor distances*.

Our next goal was to obtain micrographs of a rigid molecule that had been cluster labeled on both ends. For this purpose, we prepared functionalized clusters like **2** taking advantage of the selective monosubstitution of phosphine for iodide,⁴ not CO. The clusters were spaced apart by using synthetic methods that we have previously developed⁶ for the preparation of long, yet soluble, molecular lines (Scheme I). The resulting compound **3**⁷ has a rigid, linear spacer of 28 Å and flexible COCH₂CH₂ connectors. Importantly, **3** was soluble in organic solvents allowing purification, reliable assays of purity and structure, and application as a CHCl₃ solution to the carbon film substrate used for STEM. As anticipated, STEM micrographs (Figure 1A) showed paired 6-Å spots corresponding to individual molecules of **3**. The separations between 20 pairs were 20–37 Å (\bar{x} = 27 ± 4 Å). This range covers virtually all of the conformations available to the flexible con-

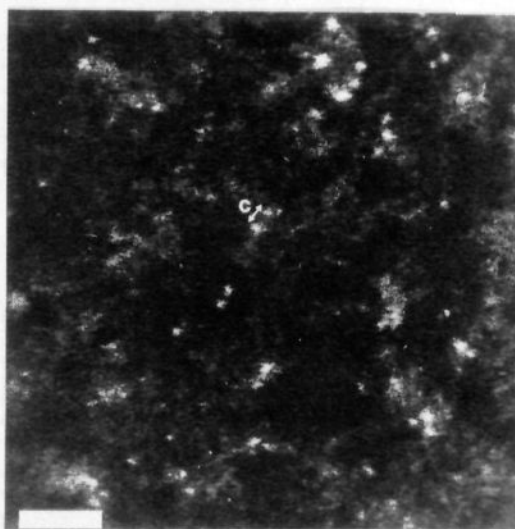
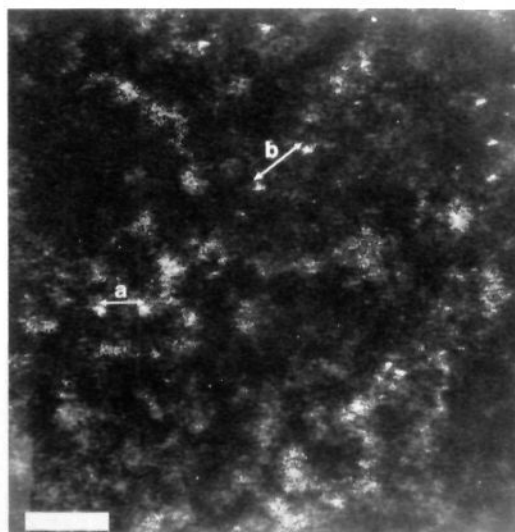


Figure 1. Dark field STEM images of **3** (A) (top) and **5** (B) (bottom). Dose 3.2 × 10⁵ e/nm²: (a) 26 Å, (b) 38 Å, (c) 10 Å. Bar = 50 Å.

nectors and shows that individual molecules adopt a variety of conformations on the surface. More accurate distance measurements between rigidly placed sites on organic or biological molecules will require less flexible attachments than any of those previously employed.^{2,3}

A shorter bis cluster compound **5**⁸ also gave 6-Å, paired spots (Figure 1B). The clusters were stationary in successive scans and did not damage substantially. Analysis of two typical micrographs containing 66 spots showed that 62 of them were correlated as pairs. The average separation for 100 pairs was 13 ± 4 Å. This affirms that the distances measured for **3** were not artifactual.

In summary, we have shown that small Ir₄(CO)₁₁ clusters are useful for STEM labelling; we have synthesized a linear spacer, attached the cluster to each end, and successfully correlated the cluster separation distances to those expected from molecular models. We expect that cluster labeling will allow us to measure the length of molecular lines on a surface, a measurement not currently feasible.

(8) Compound **5** (4,4'-dihydroxybiphenyl plus Cl(C₆H₅)₂P, followed by reaction of the phosphinite product with **1** gave **5**): ¹H NMR (CDCl₃) δ 7.7–7.6 (m, 8 H), δ 7.6–7.4 (7, 12 H), δ 7.2–7.1 (m, 4 H), δ 6.8–6.7 (m, 4 H); IR (THF) ν_{CO} 2088 m, 2056 vs, 2035 m sh, 2020 s, 1847 m, 1826 m, cm⁻²; MS, FAB calcd (M + 1) (²⁹²Ir₂, ²⁹³Ir₄) 2709.7, obsd 2709.2; isotope cluster mass (M + 1) calcd 2706 (20), 2707 (69), 2708 (43), 2709 (100), 2710 (60), 2711 (93), 2712 (53), 2713 (53), obsd 2706 (32), 2707 (66) 2708 (73), 2709 (100), 2710 (76), 2711 (98), 2712 (56), 2713 (50).

(4) Unpublished work of Roesslet and Gladfelter (Roesslet, K.; Gladfelter, W. University of Minnesota). Ros, R.; et al. *J. Chem. Soc., Dalton Trans.* **1986**, 2411.

(5) Mass analysis used the system described by Wall and Hainfeld (Wall, J. S.; Hainfeld, J. F.) *Ann. Rev. Biophys., Biophys. Chem.* **1986**, *15*, 355. Due to the high magnification (2 × 10⁶), an extrapolated mass-per-unit-intensity calibration factor from tobacco mosaic virus was used. The radius of integration of 5 Å was chosen to account for possible misplacement of the center of integration.

(6) Christophel, W. C.; Miller, L. L. *J. Org. Chem.* **1986**, *51*, 4169. Kenny, P. W.; Miller, L. L.; Chiba, T. *J. Org. Chem.* **1987**, *52*, 4327.

(7) Compound **3** (purified by preparative TLC on SiO₂ and then crystallization from toluene/CH₂Cl₂): ¹H NMR (CDCl₃) δ 8.96 (s, 4 H), 7.62–7.15 (m, 4 H), 2.91–2.77 (m, 4 H), 2.50–2.36 (m, 4 H), 1.43 (s, 36 H); IR (KBr) ν_{CO} 2089, 2053, 2014, 1885, 1844, 1818, 177 average mass of the isotope envelope; C₁₃₀H₉₄Ir₈N₄O₃₀P₂; calcd (m + 1) 3791.9, obsd 3792.3.

